Photoinduced Oxidation of Tryptamine Derivatives. Formation of Pyrrolo[2,3-b]indole and N^b -(4-Cyanobutadienyl)tryptamine

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Summary Irradiation (253·7 nm) of N^aN^b -dimethyltryptamine with pyridine N-oxide produced 1,8-dimethyl-3a-hydroxy-2,3,3a,8a-tetrahydropyrrolo[2,3-b]indole (4), whilst with visible light N^b -(4-cyanobutadienyl)- N^aN^b -dimethyltryptamine (5) was obtained.

RECENTLY, considerable interest has arisen concerning the mechanism of biological oxidations of aromatic substrates which occur concomitant with an NIH shift; arene oxides have been shown to be intermediates.¹

The photolysis of pyridine N-oxide in the presence of aromatic substrates produces arene oxides directly and

serves as a useful model reaction for such enzymatic oxidations.² A good model for the dioxygenase-type³ oxidation of tryptphan to kynuenine derivatives is provided by the photo-oxidation reaction.⁴ The frequent occurrence in nature, however, of tryptophan derivatives oxidized in the benzene ring, e.g. 5-hydroxytryptophan,³ serotonin,³ dehydrobufotenine⁵ and the sporidesmins⁶ indicate that oxidative pathways other than the cleavage of the pyrrole ring 2,3-bond are operative and encourages the search for the appropriate model reactions.

We report that the oxidation of N^aN^b -dimethyltryptamine (1) in CH_2Cl_2 , by photolysing (253.7 nm) pyridine

N-oxide (2), produces after 1 h, 1,8-dimethyl-3a-hydroxy-2,3,3a,8a-tetrahydropyrrolo[2,3-b]indole (4) (10%). Anhydro analogues of (4) have been reported, and have been converted into 3a-hydroxypyrroloindoles. Our one-step production of this structure, with a hydroxyl group in the same position as found in the sporidesmins, probably proceeds by participation of the ethylamine side chain in

opening an intermediate 2,3-oxide (3). (4); λ_{max} (EtOH) 250 (ϵ 6500), 298 nm (2360), λ_{max} (EtOH-HCl) 241 (ϵ 6100), 292 nm (2560); 8 $v_{\rm max}$ (CHCl₃) 3650, 3340 cm⁻¹ (OH); m/e204 (M+, 96%); δ (CDCl₃) 2·14—2·33 (m, -CH₂), 2·52 (s, $N^{b}-Me$), 2.68-2.90 (m, $N-CH_{2}$), 2.90 (s, $N^{a}-Me$), 4.25(s, N-CH-N), 4.44 p.p.m. (broad s, OH).

However, Nb-(4-cyanobutadienyl)tryptamine (5) was obtained in 20% yield, with only traces of (4), when a mixture of (1) and (2) was irradiated with visible light (200 W), in the presence or absence of air, providing new evidence for 1,2-oxazepine (8) or its ring opened tautomer 4-cyanobut-4-enal (9) as intermediates in the photolysis of (2). (5) was rapidly destroyed when irradiated in CH₂Cl₂ at 253.7 nm. (5); λ_{max} 225 (ϵ 35,000), 339 nm (30,400); v_{max} (CHCl₃) 2195 (C \equiv N), 1622 cm⁻¹ (C=C); δ (CDCl₃) 2.78, 2.85 (trans, cis Nb-Me), 3.73 (s, Na-Me), 4.31-5.52, 6.42-6.77 (m, vinylic H), 6.81 p.p.m. (s, α -H); m/e 265 $(M^{+}).$

Hydrogenation of (5) (Pd-C) led to the uptake of hydrogen (2 mol) and gave the saturated nitrile (6); ν_{max} (CHCl₃) 2245 cm⁻¹ (C \equiv N); m/e, 269 (M⁺). The irradiation of Nmethylaniline and (2) with visible light gave (7) in 25% yield. The structure of which was proved by i.r., n.m.r., u.v., and mass spectral data.

2-Formylpyrrole is known to arise from the photoisomerization of (2), in which an intermediate 1,2-oxazepine (8) was suggested without direct evidence. We suggest that (1) effects a base-catalysed opening of (8) in a manner analogous to the known base-catalysed ring opening of isoxazoles¹⁰ giving (9), which then reacts with (1) to give (5). Supporting this mechanism, is the isolation of photoisomer analogous to (9), viz., β -formamidoacrylonitrile (10), by irradiation of pyrimidine N-oxide. 11

Further studies on the scope of this reaction are in

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- ¹ J. W. Daly, D. M. Jerina, and B. Witkop, Arch. Biochem. Biophys., 1968, 128, 517; D. M. Jerina, J. W. Daly, B. Witkop, P. Zaltman-Nirenberg, and S. Udenfriend, J. Amer. Chem. Soc., 1968, 90, 6525.

 ² D. M. Jerina, D. R. Boyd, and J. W. Daly, Tetrahedron Letters, 1970, 457.
 - ³ O. Hayaishi and M. Nozaki, Science, 1969, 164, 398.
- W. E. Savige, Austral. J. Chem., 1971, 24, 1285 and references therein.
 F. Marki, A. V. Robertson, and B. Witkop, J. Amer. Chem. Soc., 1961, 83, 3341.
 J. W. Ronaldson, A. Taylor, E. P. White, and R. J. Abraham, J. Chem. Soc., 1963, 3172.
 M. Ohno, T. F. Spande, and B. Witkop, J. Amer. Chem. Soc., 1970, 92, 343.
 H. F. Hodson and G. F. Smith, J. Chem. Soc., 1957, 1877.
 P. L. Kumler and O. Buchardt, Chem. Comm., 1968, 1321; J. Streith and C. Sigwalt, Tetrahedron Letters, 1966, 1347; M. Ishikawa, Vernella J. Velage and Soc. Vernella Tetrahedron 1969, 1321; J. Streith and C. Sigwalt, Tetrahedron Letters, 1966, 1347; M. Ishikawa, C. Kaneko, I. Yokoe, and Sa. Yamada, Tetrahedron, 1969, 25, 295.
 - 10 N. K. Kochetkov and S. D. Sokolov, Adv. Heterocyclic Chem., 1963, 365.
- ¹¹ J. Streith and P. Martz, Tetrahedron Letters, 1969, 4899.